www.nature.com/cgt

REVIEW Molecular imaging and cancer gene therapy

Z Saadatpour¹, G Bjorklund², S Chirumbolo³, M Alimohammadi⁴, H Ehsani⁵, H Ebrahiminejad⁶, H Pourghadamyari⁷, B Baghaei⁸, HR Mirzaei⁹, A Sahebkar¹⁰, H Mirzaei¹¹ and M Keshavarzi¹²

Gene therapy is known as one of the most advanced approaches for therapeutic prospects ranging from tackling genetic diseases to combating cancer. In this approach, different viral and nonviral vector systems such as retrovirus, lentivirus, plasmid and transposon have been designed and employed. These vector systems are designed to target different therapeutic genes in various tissues and cells such as tumor cells. Therefore, detection of the vectors containing therapeutic genes and monitoring of response to the treatment are the main issues that are commonly faced by researchers. Imaging techniques have been critical in guiding physicians in the more accurate and precise diagnosis and monitoring of cancer patients in different phases of malignancies. Imaging techniques such as positron emission tomography (PET) and single-photon emission computed tomography (SPECT) are non-invasive and powerful tools for monitoring of the distribution of transgene expression over time and assessing patients who have received therapeutic genes. Here, we discuss most recent advances in cancer gene therapy and molecular approaches as well as imaging techniques that are utilized to detect cancer gene therapeutics and to monitor the patients' response to these therapies worldwide, particularly in Iranian Academic Medical Centers and Hospitals.

Cancer Gene Therapy advance online publication, 18 November 2016; doi:10.1038/cgt.2016.62

INTRODUCTION

Cancer is a huge health concern worldwide, and therefore several molecular therapeutic approaches have been recently developed,¹⁻⁶ yet with many difficulties and progressing issues. Many new therapeutic attempts to combat cancer have been made so far, many of which focus on gene therapeutic strategies.^{5,8,9} It has been recently shown that cellular gene therapies may be considered as a new and promising platform for cancer therapy. However, gene and cell therapies have been associated with some limitations.^{10,11} One of these limitations is the difficulty in tracking therapeutic genes and cells in patients' body as well as monitoring of patients' response to the treatment. In this regard, molecular imaging could be a useful tool for tracking and monitoring of cell and gene therapies for cancer and following up of the treatment outcome.^{10–12} In a more general way, these techniques enable us to monitor biological processes at cellular and subcellular levels in living organisms and might lead to various applications in other human diseases besides cancer.¹² The role of imaging in cancer therapy, particularly in cancer gene therapy, has also raised some controversies, much of which are coming from technical concerns. Imaging may bridge the gap between cell and/or gene therapies and health outcomes by elucidating the mechanisms of action through longitudinal monitoring.^{10,11}

Imaging-guided delivery of gene-targeting therapeutics represents a robust tool in the fight against cancer through gene transfer or RNA interference.¹ An imaging-driven approach should allow, for example, the translating RNA interference cancer therapy into the clinical practice, as the technique could address many issues related to administration route, systemic blood circulation and cellular or tissue barriers, besides safety and ethical concerns.¹ Although cancer in Iran and Arabic countries still has very low survival rate,^{2,13} gene therapy may represent an innovative and promising therapeutic strategy for cancer treatment, though technical concerns regarding its feasibility remain to be addressed.¹⁴

IMAGING IN GENE TARGETING AND THERAPY: WHEAT AND CHAFFS

Cancer is one of the main public health problems worldwide.¹² Various studies assessed different dimensions of this disease. To date, through deepening understanding of molecular/cellular pathways involved in different cancers, researchers have designed many therapies and therapeutic regimens for cancer, although cancer yet remains as a main problem in numerous communities.^{10–12} Among various current anticancer therapies, gene therapy has emerged as a new platform for the cancer treatment.^{10–12} Gene therapy enables researchers to introduce a new therapeutic gene in receipt cells by specific vector systems such as plasmid, transposon and lentivirus. One of the major challenges in these systems is how to detect these vectors in

E-mail: maryamkeshavarzi2@gmail.com

Received 6 September 2016; revised 21 September 2016; accepted 23 September 2016

¹Bozorgmehr Imaging Center, Isfahan University of Medical Sciences, Isfahan, Iran; ²Council for Nutritional and Environmental Medicine, Mo i Rana, Norway; ³Department of Neurological and Movement Sciences, University of Verona, Verona, Italy; ⁴Department of Oral and Maxillofacial Radiology, School of Dentistry, Mazandaran University of Medical Sciences, Sari, Iran; ⁵Department of Periodontology, School of Dentistry, Mazandaran University of Medical Sciences, Sari, Iran; ⁶Department of Oral and Maxillofacial Radiology, School of Dentistry, Mazandaran University of Medical Sciences, Sari, Iran; ⁶Department of Periodontology, School of Dentistry, Mazandaran University of Medical Sciences, Sari, Iran; ⁶Department of Oral and Maxillofacial Radiology, School of Dentistry, Rerman University of Medical Sciences, Kerman, Iran; ⁷Department of Biochemistry, School of Medicine, Tehran University of Medical Sciences, Tehran, Iran; ⁸Department of Endodontics, School of Dentistry, Rafsanjan University of Medical Sciences, Rafsanjan, Iran; ⁹Department of Immunology, School of Medicine, Tehran University of Medical Sciences, Nashhad, Iran; ¹⁰Biotechnology Research Center, Mashhad University of Medical Sciences, Mashhad, Iran; ¹¹Department of Medical Sciences, Khorramabad, Iran. Correspondence: Dr M Keshavarzi, Department of Oral and Maxillofacial Radiology, School of Dentistry, Lorestan University of Medical Sciences, Khorramabad, Iran.

Molecular imaging and cancer gene therapy Z Saadatpour *et al*

2

Imaging technique	Advantage (s)	Disadvantage (s)
PET	High sensitivity	High costs
	High penetration depth	
SPECT	Different radioisotopes	Influence on biomolecule
	High penetration depth	Limitation in spatial resolution
	Low costs	
	Labeling simultaneously	
MRI	High spatial resolution	Time consuming
	Extracting physiological, molecular and anatomical information	High costs
Computed tomography	High spatial resolution	Limitation in biocompatible contrast
	5	Radiation exposure
		Limited imaging in soft tissue
Ultrasound	Clinical friendly	Limitation in bone and lung
	High spatial and temporal resolution	J
	Low costs	
Optical	High temporal resolution	Low penetration of tissue
	Non-invasive functional imaging	Non-quantification
	High sensitivity	·
	Low costs	

Imaging technique	Main type of cancer	Markers	References
X-ray-based approaches			
Flat panel CR	Bone		43
Digital radiography, TAC	Lung		44
Multislice CT system	Bone		45
Magnetic resonance imaging (MRI)	Breast cancer		46
Diffusion waived imaging	Neuroendocrine and hepatic metastasis		47
MR elastography	Breast	A	48
MR perfusion imaging	Breast	В	49
MR spectroscopy	Breast		50
MR-guided focused ultrasound	HCC		51
MR-guided galvanotherapy	Prostate		52
Sonography–ultrasound			
Miniaturization of ultrasound systems	Prostate		53–55
Acoustic radiation force impulse imaging	Hepatic malignancies		
High-intensity-focused ultrasound	Prostate		
Non-ionizing electromagnetic imaging			
Photo- and thermoacoustic imaging	Breast		56–58
Electrical impedance tomography			
Near-infrared optical tomography			
Nuclear medicine imaging			50.50
PET	Colon	C	59,60
SPECT		D	

recycling and expression.

preclinical and clinical studies. Imaging is home to a wide array of clinical diagnostic and therapeutic technologies.^{5,8,9} In regenerative medicine, imaging not only is greatly useful in unveiling damaged tissues but also in monitoring of safety and efficacy of a defined targeted therapy (Table 1).¹⁵ Table 2 lists the main imaging techniques used in cancer diagnosis and therapy.

Positron emission tomography (PET) with I-124-labeled 2'-fluoro-2'-deoxy-1b-D-arabino-furanosyl-5-iodo-uracil ([¹²⁴I]-FIAU)—a specific marker substrate for gene expression of HSV-1-tk—has been recently used to detect the location, magnitude and extent of vector-mediated HSV-1-*tk* gene expression in a phase I/II clinical trial of gene therapy for recurrent glioblastoma.¹⁶ This approach has yet raised some critical issues in identification of the exogenous gene expression introduced into the patients with glioma.¹⁷ The [¹²⁴I]-FIAU substrate does not succeed in penetrating intact bloodbrain barrier (BBB); however, it appears that this substrate can penetrate in disrupted areas of BBB (for example, glioblastomas). Accordingly, to detect localization of HSV-1-*tk* gene expression in

Experimental system	Background	Objective	Reference
Cell therapy			
CSCs targeting	Target and/or modifying CSC markers	Gene target immunotherapy	61,62
MSCs	Stem cell research in cancer treatment	Cell EMT/MET transition	
Receptor inhibitors			
1	Targeting c-kit as a novel gene therapy	Gene-protein molecular targeting	62
Nanotechnology			
Polymeric nanoparticles	Nanocarriers used for gene therapy	PLGA, PEIs	63
Graphene oxide nanocarriers for siRNAs	Polymeric nanoparticles, lipid-based carriers (liposomes/micelles), dendrimers, carbon nanotubes and gold nanoparticles (nanoshells/ nanocages)	PL-PEG and CPPs	64
D3-loaded nanoparticles	Tarlocages		65
Gene therapy			5
siRNA/miRNA-interfering RNAs	siRNAs in ovarian cancer	Interfering RNA	66
Epigenetics	Epigenetics in multiple sclerosis	Perspective on the use of epigenetics	67
Transcription factors	BTB and CNC homology 1, basic leucine zipper transcription factor 1	BACH1 targeting as a new tool	68
Long non-coding RNAs	LncRNAs as targets	Targeting LncRNAs	69
LNA-modified oligonucleotides	LNA-anti-miR-21 (50 pmol μ l ⁻¹)	Used in colorectal cancer	43
In silico analysis	G-guadruplexes in WT1 oncogene	Drug design	43

Abbreviations: CPP, cell-penetrating peptide; CSC, cancer stem cell; EMT, epithelial-mesenchymal transition; LNA, locked nucleic acid; LncRNAs, long noncoding RNAs; MET, mesenchymal-epithelial transition; MSC, mesenchymal stem cell; PEI, polyethyleneimine; PLGA, poly lactic-co-glycolic acid; PL-PEG, phospholipid-based amphiphilic polymer; siRNA, short interfering RNA.

the central nervous system by FIAU PET, washout of nonspecific radioactivity for several days was highly recommended.¹⁸ PET technique, as either micro-PET or micro-computed tomography, is also currently included in gene therapy protocol for small animal models.^{19,20} By emerging more sensitive and high-capacity nuclear-imaging methods, Zinn *et al.*²¹ demonstrated that utilizing singlephoton emission computed tomography (SPECT) methods such as gamma camera imaging provided a high-capacity method to image gene transfer to cancer cells. They showed that gamma camera imaging can detect the expression of the hSSTr2 reporter protein in human cancer cells (like A-427 non-small cell lung, SKOV3.ip1 ovarian, MDA-MB-468 breast and BxPC-3 pancreatic) by imaging an internalized [sup 99 m]Tc-labeled, hSSTr2-binding peptide. The approach was later developed as radiolabeled imaging, for example, by inducing tumor cells to express the human somatostatin receptor subtype 2 (hSSTr2)—a high-affinity receptor for radiolabeled somatostatin analogs.²² Although through improvement of imaging technology, PET and SPECT methods have notably improved the ability to detect and to follow up gene transfer in tumor cells, some barriers including high cost, timeconsuming nature, technique difficulty, reliance on radioactive reagents, immature reimbursement structure and the need for continuous training negatively influenced on their wide adoption in clinical practice.^{23,24} Magnetic resonance imaging (MRI) represents a more available alternative.^{25,26} In gene silencing using short interfering RNA (siRNA), the complexation of siRNA with polyethyleneimine prevents rapid degradation of siRNA in tumors and, more interestingly, enhances gene transfection efficiency.^{26,27} Interestingly, PEGylated polyethyleneimine modified with gadolinium chelator (DOTA), which has been used as a template for synthesis of polyethyleneimine/gadolinium-related nanoparticles, is a well-known nanocarrier for gene silencing and has potential application in the MRI ambient.²⁸ The use of MRI in gene silencing and gene transfer has been adopted in recent years;²⁹⁻³¹ however,

its application has been expanded into not only oncology but also several other fields. MRI is a much more widely used and accepted method than PET, SPECT and CT. Combined with new techniques such as cine-MRI and functional methods such as perfusion- and diffusion-weighted imaging, MRI may also be considered as an alternative tool to conventional radiological-based approaches such as radiography, computed tomography and PET/CT imaging in the evaluation of cancer patients.³² Some molecules, such as creatine kinase reporter gene, act as sensitive bio-magnetic detectors in MRI for gene transfer, providing advantages for utilizing MRI in gene transfer.³³ CK is particularly abundant in the human muscle and the brain, but not in the liver; therefore, combined application of CK reporter gene and the phosphorus-31 magnetic resonance spectroscopy is particularly useful to detect and monitor any liver-directed gene transfer.³³ Emerging of new imaging approaches, such as quantitative three-dimensional oxygen imaging using electron paramagnetic resonance imaging and hyperpolarized ¹³C metabolic MRI, might be additionally useful in monitoring and prediction of tumor response to anticancer gene therapy through tracing reporter gene-derived new metabolic reactions.34 The evolution of new MRI probes may extend the application of this imaging technique to cancer gene therapy; for example, in prostate cancer, introduction of hyperpolarized ¹³C-organic acids, such as lactic acid, pyruvic acid or even alanine and most recently hyperpolarized [1-¹³C]pyruvate, has enhanced sensitivity of conventional MRI.^{35,36}

In a more general way, imaging in gene therapy has been widely adopted both in MRI with ancillary approaches to improve the MRI resolution and more recently in positron-emitting techniques. Further methods are currently being developed to monitor cancer gene therapy outcome. For example, in a spontaneous tumor-bearing animal model, intratumoral interleukin-12 electrogene therapy was monitored via contrast-enhanced ultrasound.³⁷ In another study, Aalinkeel *et al.*³⁸ utilized MRI and ultrasonography to monitor tumor perfusion following

cationic polylactide (CPLA) interleukin-8 siRNA nanocomplexes.³⁸ Various studies have reported that, imaging techniques such as PET scan, CT or ultrasound (for example, ultrasound-guided endoscopy), combined by other technologies, can be used for monitoring, management and differential diagnosis of tumors, particularly prostate cancer.³⁹

The identification of various markers in response to different therapies could contribute to better treatment of cancer.^{40,70–74} Many of these molecules and agents could be used as markers for assessment of response to treatment in various cancers. For example, in tumor cells, there is an increase in glycolysis rate, DNA synthesis and angiogenesis—the processes that could be utilized for monitoring of cancer patients. Various studies revealed that there are two classes of imaging techniques associated with gene therapy including biodistribution and transduction imaging. Transduction imaging methods could detect transgene-mediated protein production.¹⁰ On the other hand, biodistribution-imaging methods could track gene delivery vectors.⁴¹ The individual using the above-mentioned imaging techniques is associated with some limitations. For example, transduction pattern may show an inaccurate image of viral biodistribution so that a virus could enter into several cells, but its transgene of interest is not expressed in all transduced cells. Hence, one of the main issues that should be assessed is that of transgene expression and viral particle kinetics in vivo.42 Imaging technologies use different forms of energy when interacting with different tissues. It has been shown that some techniques, including MRI and CT, are relied on energytissue interactions. On the other hand, other techniques such as SPECT and PET are relied on injection of reporter probes, as previously indicated. Imaging techniques are attempted to save time and costs through minimizing the utility of laboratory animals and time-consuming invasive techniques.¹⁰ Table 3 summarizes some of the most recent advances in gene therapy and molecular approaches that are utilized to fight cancer in Iranian Academic Medical Centers and Hospitals. In order to drive chemopreventive nanocarriers or nanostructured siRNA or microRNA (miRNA) vehicles, imaging techniques should be performed.¹

CONCLUSION

What is the best and currently available imaging approach to monitor safety, efficacy and success of a gene therapy protocol? This still represents a major concern in many countries where advanced imaging approaches and equipment are currently introduced into preclinical and clinical practices. Important considerations in choosing the most appropriate technique include gene therapy protocol, tumor type and location, safety, availability, and more precise and accurate data acquisition. Technological progress in imaging techniques will help us not only to develop safe and efficient gene therapy protocols for clinical application but also enable us to monitor therapeutic effects in cancer patients who have received therapeutic genes.

CONFLICT OF INTEREST

The authors declare no conflict of interest.

REFERENCES

- Wang J, Mi P, Lin G, Wáng YXJ, Liu G, Chen X. Imaging-guided delivery of RNAi for anticancer treatment. Adv Drug Deliv Rev 2016; 104: 44–60.
- 2 Veisani Y, Delpisheh A. Survival rate of gastric cancer in Iran; a systematic review and meta-analysis. *Gastroenterol Hepatol Bed Bench* 2016; **9**: 78.
- 3 Mirzaei H, Gholamin S, Shahidsales S, Sahebkar A, Jafaari MR, Mirzaei HR *et al.* MicroRNAs as potential diagnostic and prognostic biomarkers in melanoma. *Eur J Cancer* 2016; **53**: 25–32.

- 4 Faghihloo E, Araei Y, Mohammadi M, Mirzaei H, Mohammadi H, Mokhtari-Azad T. The effect of oxamflatin on the E-cadherin expression in gastric cancer cell line. *Cancer Gene Therapy* 2016; **23**: 396–399.
- 5 Mirzaei H, Sahebkar A, Yazdi F, Salehi H, Jafari M, Namdar A et al. Circulating microRNAs in hepatocellular carcinoma: potential diagnostic and prognostic biomarkers. *Curr Pharm Des* 2016 (e-pub ahead of print).
- 6 Simonian M, Mosallayi M, Mirzaei H. Circulating miR-21 as novel biomarker in gastric cancer: diagnostic and prognostic biomarker. J Cancer Res Ther 2016.
- 7 Salarini R, Sahebkar A, Mirzaei H, Jaafari M, Riahi M, Hadjati J et al. Epi-drugs and Epi-miRs: moving beyond current cancer therapies. *Curr Cancer Drug Targets* 2016; 16: 773–788.
- 8 Mirzaei HR, Sahebkar A, Salehi R, Nahand JS, Karimi E, Jaafari MR *et al.* Boron neutron capture therapy: moving toward targeted cancer therapy. 2016.
- 9 Mirzaei H, Sahebkar A, Jaafari M, Hadjati J, Javanmard S, Mirzaei H et al. PiggyBac as a novel vector in cancer gene therapy: current perspective. *Cancer Gene Ther* 2016; 23: 45–47.
- 10 Shah K, Jacobs A, Breakefield X, Weissleder R. Molecular imaging of gene therapy for cancer. *Gene Ther* 2004; **11**: 1175–1187.
- 11 Sato T, Liu X, Nelson A, Nakanishi M, Kanaji N, Wang X et al. Reduced miR-146a increases prostaglandin E2 in chronic obstructive pulmonary disease fibroblasts. Am J Respir Crit Care Med 2010; 182: 1020–1029.
- 12 Räty JK, Liimatainen T, Kaikkonen MU, Gröhn O, Airenne KJ, Ylä-Herttuala S. Non-invasive imaging in gene therapy. *Mol Ther* 2007; **15**: 1579–1586.
- 13 Rahou BH, El Rhazi K, Ouasmani F, Nejjari C, Bekkali R, Montazeri A et al. Quality of life in Arab women with breast cancer: a review of the literature. Health Quality Life Outcomes 2016; 14: 1.
- 14 Borna H, Imani S, Iman M, Azimzadeh Jamalkandi S. Therapeutic face of RNAi: in vivo challenges. Exp Opin Biol Ther 2015; **15**: 269–285.
- 15 Naumova AV, Modo M, Moore A, Murry CE, Frank JA. Clinical imaging in regenerative medicine. *Nat Biotechnol* 2014; **32**: 804–818.
- 16 Jacobs A, Voges J, Reszka R, Lercher M, Gossmann A, Kracht L et al. Positronemission tomography of vector-mediated gene expression in gene therapy for gliomas. Lancet 2001; 358: 727–729.
- 17 Ia Fougère C, Suchorska B, Bartenstein P, Kreth F-W, Tonn J-C. Molecular imaging of gliomas with PET: opportunities and limitations. *Neuro-oncology* 2011; 13: 806–819.
- 18 Jacobs A, Bräunlich I, Graf R, Lercher M, Sakaki T, Voges J et al. Quantitative kinetics of [124] FIAU in cat and man. J Nucl Med 2001; 42: 467–475.
- 19 Xu H, Guo R, Jin Y, Li B. [Gene-targeted radiation therapy mediated by radiationsensitive promoter in lung adenocarcinoma and the feasibility of micro-PET/CT in evaluation of therapeutic effectiveness in small animals]. *Zhongua Zhong Liu Za Zhi* 2014; **36**: 329–334.
- 20 Collins S, Hiraoka K, Inagaki A, Kasahara N, Tangney M, PET imaging for gene & cell therapy. *Curr Gene Ther* 2012; **12**: 20–32.
- 21 Zinn K, Chaudhuri T, Buchsbaum D, Mountz J, Rogers B. Detection and measurement of *in vitro* gene transfer by gamma camera imaging. *Gene Ther* 2001; 8: 4.
- 22 Buchsbaum DJ, Chaudhuri TR, Yamamoto M, Zemn KR. Gene expression imaging with radiolabeled peptides. Ann Nucl Med 2004; 18: 275–283.
- 23 Bateman TM. Advantages and disadvantages of PET and SPECT in a busy clinical practice. J Nucl Cardiol 2012; 19: 3–11.
- 24 Simonova M, Wall A, Weissleder R, Bogdanov A. Tyrosinase mutants are capable of prodrug activation in transfected nonmelanotic cells. *Cancer Res* 2000; 60: 6656–6662.
- 25 Moore A, Josephson L, Bhorade RM, Basilion JP, Weissleder R. Human transferrin receptor gene as a marker gene for MR imaging 1. *Radiology* 2001; **221**: 244–250.
- 26 Xie L, Tan Y, Wang Z, Liu H, Zhang N, Zou C et al. Epsilon-caprolactone modified polyethyleneimine as efficient nanocarriers for siRNA delivery in vivo. ACS Appl Mater Interfaces 2016 (e-pub ahead of print).
- 27 Aigner A. Gene silencing through RNA interference (RNAi) *in vivo* strategies based on the direct application of siRNAs. *J Biotechnol* 2006; **124**: 12–25.
- 28 Zhou B, Xiong Z, Zhu J, Shen M, Tang G, Peng C et al. PEGylated polyethylenimine-entrapped gold nanoparticles loaded with gadolinium for dual-mode CT/MR imaging applications. *Nanomedicine* 2016; 11: 1639–1652.
- 29 Ichikawa T, Högemann D, Saeki Y, Tyminski E, Terada K, Weissleder R *et al.* MRI of transgene expression: correlation to therapeutic gene expression. *Neoplasia* 2002; 4: 523–530.
- 30 Kircher MF, Allport JR, Graves EE, Love V, Josephson L, Lichtman AH et al. In vivo high resolution three-dimensional imaging of antigen-specific cytotoxic T-lymphocyte trafficking to tumors. Cancer Res 2003; 63: 6838–6846.
- 31 Tavri S, Jha P, Meier R, Henning TD, Müller T, Hostetter D et al. Optical imaging of cellular immunotherapy against prostate cancer. *Mol Imaging* 2009; 8: 1.

- 32 Guimaraes MD, Hochhegger B, Santos MK, Santana PRP, Sousa Júnior AS, Souza LS et al. Magnetic resonance imaging of the chest in the evaluation of cancer patients: state of the art. Radiol Bras 2015; 48: 33–42.
- 33 Li Z, Qiao H, Lebherz C, Choi SR, Zhou X, Gao G et al. Creatine kinase, a magnetic resonance-detectable marker gene for quantification of liver-directed gene transfer. Hum Gene Ther 2005; 16: 1429–1438.
- 34 Matsumoto S. Prediction of cancer treatment response by physiologic and metabolic imaging. *Yakugaku Zasshi* 2016; **136**: 1101.
- 35 Albers MJ, Bok R, Chen AP, Cunningham CH, Zierhut ML, Zhang VY et al. Hyperpolarized 13C lactate, pyruvate, and alanine: noninvasive biomarkers for prostate cancer detection and grading. *Cancer Res* 2008; 68: 8607–8615.
- 36 Nelson SJ, Kurhanewicz J, Vigneron DB, Larson PE, Harzstark AL, Ferrone M et al. Metabolic imaging of patients with prostate cancer using hyperpolarized [1-13C] pyruvate. Sci Transl Med 2013; 5: 198ra08–ra08.
- 37 Cicchelero L, Denies S, Haers H, Vanderperren K, Stock E, Van Brantegem L et al. Intratumoural interleukin 12 gene therapy stimulates the immune system and decreases angiogenesis in dogs with spontaneous cancer. Vet Comp Oncol 2016.
- 38 Aalinkeel R, Nair B, Chen CK, Mahajan SD, Reynolds JL, Zhang H et al. Nanotherapy silencing the interleukin-8 gene produces regression of prostate cancer by inhibition of angiogenesis. *Immunology* 2016; **148**: 387–406.
- 39 Bournet B, Buscail C, Muscari F, Cordelier P, Buscail L. Targeting KRAS for diagnosis, prognosis, and treatment of pancreatic cancer: Hopes and realities. *Eur J Cancer* 2016; 54: 75–83.
- 40 Brindle K. New approaches for imaging tumour responses to treatment. *Nat Rev Cancer* 2008; **8**: 94–107.
- 41 Bogdanov A, Weissleder R. The development of *in vivo* imaging systems to study gene expression. *Trends Biotechnol* 1998; **16**: 5–10.
- 42 Massoud TF, Gambhir SS. Molecular imaging in living subjects: seeing fundamental biological processes in a new light. *Genes Dev* 2003; 17: 545–580.
- 43 Nedaeinia R, Sharifi M, Avan A, Kazemi M, Rafiee L, Ghayour-Mobarhan M et al. Locked nucleic acid anti-miR-21 inhibits cell growth and invasive behaviors of a colorectal adenocarcinoma cell line: LNA-anti-miR as a novel approach. Cancer Gene Ther 2016; 23: 246–253.
- 44 Campadelli P, Casiraghi E, Artioli D. A fully automated method for lung nodule detection from postero-anterior chest radiographs. *IEEE Trans Med Imaging* 2006; 25: 1588–1603.
- 45 Simon CJ, Dupuy DE (eds). Seminars in Musculoskeletal Radiology. Thieme Medical Publishers Inc.: New York, NY, USA: 2006.
- 46 Bhattacharyya M, Ryan D, Carpenter R, Vinnicombe S, Gallagher C. Using MRI to plan breast-conserving surgery following neoadjuvant chemotherapy for early breast cancer. Br J Cancer 2008; 98: 289–293.
- 47 Liapi E, Geschwind J-F, Vossen JA, Buijs M, Georgiades CS, Bluemke DA et al. Functional MRI evaluation of tumor response in patients with neuroendocrine hepatic metastasis treated with transcatheter arterial chemoembolization. Am J Roentgenol 2008; **190**: 67–73.
- 48 Plewes DB, Bishop J, Samani A, Sciarretta J. Visualization and quantification of breast cancer biomechanical properties with magnetic resonance elastography. *Phys Med Biol* 2000; **45**: 1591.
- 49 Helbich TH, Roberts TP, Gossmann A, Wendland MF, Shames DM, Adachi M et al. Quantitative gadopentetate-enhanced MRI of breast tumors: testing of different analytic methods. *Magn Reson Med* 2000; 44: 915–924.
- 50 Bartella L, Huang W. Proton (1H) MR spectroscopy of the breast 1. *Radiographics* 2007; 27: S241–S252.
- 51 Kopelman D, Inbar Y, Hanannel A, Dank G, Freundlich D, Perel A et al. Magnetic resonance-guided focused ultrasound surgery (MRgFUS). Four ablation treatments of a single canine hepatocellular adenoma. HPB 2006; 8: 292–298.
- 52 Vogl TJ, Mayer HP, Zangos S, Selby JB Jr, Ackermann H, Mayer FB. Prostate Cancer: MR imaging–guided galvanotherapy—technical development and first clinical results 1. *Radiology* 2007; 245: 895–902.
- 53 Narayan P, Gajendran V, Taylor SP, Tewari A, Presti JC, Leidich R et al. The role of transrectal ultrasound-guided biopsy-based staging, preoperative serum prostate-specific antigen, and biopsy Gleason score in prediction of final pathologic diagnosis in prostate cancer. Urology 1995; 46: 205–212.

- 54 Fahey B, Nelson R, Bradway D, Hsu S, Dumont D, Trahey G. In vivo visualization of abdominal malignancies with acoustic radiation force elastography. *Phys Med Biol* 2007; **53**: 279.
- 55 Paliwal S, Sundaram J, Mitragotri S. Induction of cancer-specific cytotoxicity towards human prostate and skin cells using quercetin and ultrasound. *Br J Cancer* 2005; **92**: 499–502.
- 56 Pogue BW, Poplack SP, McBride TO, Wells WA, Osterman KS, Osterberg UL *et al.* Quantitative hemoglobin tomography with diffuse near-infrared spectroscopy: pilot results in the breast 1. *Radiology* 2001; **218**: 261–266.
- 57 Kao T-J, Saulnier GJ, Xia H, Tamma C, Newell J, Isaacson D. A compensated radiolucent electrode array for combined EIT and mammography. *Physiol Meas* 2007; **28**: S291.
- 58 Ntziachristos V, Yodh A, Schnall MD, Chance B. MRI-guided diffuse optical spectroscopy of malignant and benign breast lesions. *Neoplasia* 2002; 4: 347–354.
- 59 Hung S-C, Deng W-P, Yang WK, Liu R-S, Lee C-C, Su T-C et al. Mesenchymal stem cell targeting of microscopic tumors and tumor stroma development monitored by noninvasive in vivo positron emission tomography imaging. *Clin Cancer Res* 2005; **11**: 7749–7756.
- 60 Cascini GL, Avallone A, Delrio P, Guida C, Tatangelo F, Marone P *et al.* 18 F-FDG PET is an early predictor of pathologic tumor response to preoperative radiochemotherapy in locally advanced rectal cancer. *J Nucl Med* 2006; **47**: 1241–1248.
- 61 Ranji P, Kesejini TS, Saeedikhoo S, Alizadeh AM. Targeting cancer stem cellspecific markers and/or associated signaling pathways for overcoming cancer drug resistance. *Tumor Biol* 2016 (e-pub ahead of print).
- 62 Hendijani F, Javanmard SH. Dual protective and cytotoxic benefits of mesenchymal stem cell therapy in combination with chemotherapy/radiotherapy for cancer patients. *Crit Rev Eukaryot Gene Expr* 2015; **25**: 203–207.
- 63 Bolhassani A, Javanzad S, Saleh T, Hashemi M, Aghasadeghi MR, Sadat SM. Polymeric nanoparticles: potent vectors for vaccine delivery targeting cancer and infectious diseases. *Hum Vaccin Immunother* 2014; **10**: 321–332.
- 64 Imani R, Shao W, Taherkhani S, Emami SH, Prakash S, Faghihi S. Dualfunctionalized graphene oxide for enhanced siRNA delivery to breast cancer cells. *Colloids Surf B Biointerfaces* 2016; **147**: 315–325.
- 65 Maleklou N, Allameh A, Kazemi B. Targeted delivery of vitamin D3-loaded nanoparticles to C6 glioma cell line increased resistance to doxorubicin, epirubicin, and docetaxel *in vitro*. *In Vitro Cell Dev Biol Anim* 2016 (e-pub ahead of print).
- 66 Aslani S, Jafari N, Javan MR, Karami J, Ahmadi M, Jafarnejad M. Epigenetic modifications and therapy in multiple sclerosis. *Neuromol Med* 2016 (e-pub ahead of print).
- 67 Davudian S, Mansoori B, Shajari N, Mohammadi A, Baradaran B. BACH1, the master regulator gene: a novel candidate target for cancer therapy. *Gene* 2016; 588: 30–37.
- 68 Majidinia M, Yousefi B. Long non-coding RNAs in cancer drug resistance development. DNA Rep 2016; 45: 25–33.
- 69 Malih S, Saidijam M, Malih N. A brief review on long noncoding RNAs: a new paradigm in breast cancer pathogenesis, diagnosis and therapy. *Tumor Biol* 2016; 37: 1479–1485.
- 70 Mirzaei HR, Mirzaei H, Lee SY, Hadjati J, Till BG. Prospects for chimeric antigen receptor (CAR) T cells: a potential game changer for adoptive T cell cancer immunotherapy. *Cancer Lett* 2016; **380**: 413–423.
- 71 Mirzaei H, Avan A, Salehi H, Sahebkar A, Namdar A, Mirzaei HR et al. Deciphering biological characteristics of tumorigenic subpopulations in human colorectal cancer reveals cellular plasticity. J Res Med Sci 2016; 21.
- 72 Mohammadi M, Goodarzi M, Jaafari M, Mirzaei H, Mirzaei H. Circulating microRNA: a new candidate for diagnostic biomarker in neuroblastoma. *Cancer Gene Ther* 2016; 23: 371–372.
- 73 Fathullahzadeh S, Mirzaei H, Honardoost M, Sahebkar A, Salehi M. Circulating microRNA-192 as a diagnostic biomarker in human chronic lymphocytic leukemia. *Cancer Gene Ther* 2016; 23: 327–332.
- 74 Mohammadi M, Jaafari M, Mirzaei H, Mirzaei H. Mesenchymal stem cell: a new horizon in cancer gene therapy. *Cancer Gene Ther* 2016; 23: 285–286.