Effect of royal jelly on testicular antioxidant enzymes activity, MDA level and spermatogenesis in rat experimental Varicocele model

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ABSTRACT

Varicocele is one of the most prevalent causes of infertility. It causes induction of oxidative stress, increases lipid peroxidation in the testis and disrupts spermatogenesis cycle. The aim of the study was to investigate the possible protective effects of royal jelly against varicocele induced oxidative stress, biochemical and histological alterations in the experimental varicocele model in rat. Twenty-one adult Wistar rats were divided into three groups. The control group (I), Varicocele and administration of normal saline (II), varicocele and treatment with RJ (III). At the end of the experiment, all the animals were sacrificed and testes excised. The activity of catalase (CAT), superoxide dismutase (SOD), glutathione peroxidase (GPx) and MDA levels were measured. Also, histopathological examinations, Johnsen scores and sperm parameters were determined. There was a significant increase (p < 0.05) in the activity level of CAT (0.223 ± 0.005), SOD (0.177 ± 0.0062), GPx (9.575 ± 0.318) and a significant reduction (p < 0.05) in the MDA level (2.674 ± 0.336) of the experimental varicocele treated with royal jelly when compared to the activity of CAT (0.011 ± 0.004), SOD (0.035 ± 0.0096), GPx (8.864 ± 0.397) and MDA level (4.630 ± 0.579) of the experimental varicocele and administration of normal saline. Results of the Johnsen score showed a significant increase (p < 0.05) in the mean score of the RJ group (7.94 ± 1.5) when compared to the normal saline group (6.04 ± 1.4). Therefore, RJ is a potential area for further studies and improving in spermatogenesis cycle after varicocele.

1. Introduction

Varicocele remains the most common cause of male infertility worldwide. This complication found in 15% of all men and in 19%–41% of men who present with primary infertility, representing the most common surgically treatable cause of male infertility. In men with secondary infertility, varicoceles are an underlying cause in 45%–81% of patients (Pastuszak and Wang, 2015). Ten studies in meta-analysis involving 1232 men show that varicocele is associated with a reduction in sperm count, motility and morphology (Agarwal et al., 2016). Studies have shown that varicocele occurrence is associated with excessive production of reactive oxygen species (ROS). Superoxide anion radical, hydrogen peroxide, oxygen singlet and proxy nitrite radical are highly reactive species (khatri and Juvekar, 2013). They can cause pathological damage by inducing oxidative changes in cellular lipids, proteins, and implicate sperm dysfunction (Smith et al., 2006; Bromfield et al., 2015; Di Meo et al., 2016). Therefore, the ROS level is an important parameter for assessing the treatment of varicocele-induced infertility. Although varicocelectomy can improve semen parameter (for example, sperm count, motility or morphology) and there is high-level evidence to suggest that in the varicocele patients, varicocelectomy results in improvement of outcomes. But the role of varicocele repair for the treatment of subfertile men has been questioned during the past decades (Agarwal et al., 2012; Cho and Seo, 2014). Some studies have shown that there is no beneficial effect of varicocele repair on a couple’s chance of conception (Evers and Collins, 2004; Evers et al., 2008). Therefore, Varicocele repair is not necessarily recommended for all infertile men with varicocele, and currently existing guidelines suggest some considerations for selecting candidates for surgical or radiological treatments. Reports have shown that Varicocele repair presents...
relatively few risks, which might include, buildup of fluid around the testicles (hydrocele), recurrence of varicocele, infection and damage to an artery. Therefore, considering the prevalence of varicocele in adolescents and surgical injury caused to correct the condition, the antioxidant treatments are necessary to counteract their harmful effects (Agarwal et al., 2005). The antioxidant compounds react in one-electron reactions with free radicals in vivo/in vitro and prevent oxidative damage. Therefore, the natural antioxidants not only play an important role in the prevention and adjunctive treatment of diseases but also can avoid the adverse reactions to human health. For example, study of 219 men with varicocele also reported an increase in sperm concentration and motility and improved morphology after 4 months using L-carnitine and acetyl-L-carnitine (Balercia et al., 2005). Oliva et al. examined the effect of 12 weeks of daily oral a dministration of pentoxifylline with zinc and folic acid on the semen quality of 36 men with varicocele-associated infertility in an open, uncontrolled study. After 4 weeks of treatment, the proportion of morphologically normal sperm cells was significantly increased; these changes persisted until at least 4 weeks after the end of treatment (Oliva et al., 2009).Wong et al. observed an increased sperm count and improved semen concentration in a controlled trial with 103 infertile and 107 fertile men who had taken 5 mg folic acid and 66 mg ZnSO4 per day for 6 months (Wong et al., 2002). Royal jelly is a natural substance that is considered to be one of the most important products of honeybees (Barnutiu et al., 2011). It is se-creted from the glands in the hypopharynx of worker bees. All larvae in the colony are fed with RJ regardless of sex or caste (Barnutiu et al., 2011). The chemical composition analysis shows that this natural substance contains a complex mixture of free amino acids, proteins, sugars, fatty acids (mainly10-hydroxy-2-decanonic acid), minerals (mainly iron and calcium), vitamins (mainly thiamine, niacin, ribo-flavin, and vitamins A, C, D and E), nucleic acids, acetylcholine, as-partic acid, and gelatin, sterols(Cavusoglu et al., 2009). Studies shows the antioxidant property of RJ (Nekeety et al., 2007; Nakajima et al., 2009; Silici et al., 2011; Ghanbari et al., 2015), especially when the RJ is used for induction of varicocele (Mahbobi et al., 2012). More than 12 antioxidant peptides in RJ have been identified showed strong hy-droxyl radical scavenging activity (Guo et al., 2009). It has also been suggested that organic acids present in RJ contribute to antioxidiant activity through metal chelation and increase the effect of total poly-phenols (Balkanska et al., 2017). The antioxidant activity of phenolic compounds is mainly due to their redox properties, which can play an important role in absorbing and neutralizing free radicals (Javanmardi et al., 2003). Studies have shown that RJ plays an important role in the immune system (Li et al., 2013), reduces blood pressure (Tokunaga et al., 2004) and has antitumor property (Shirzad et al., 2013). Thus, the aim of the present study was to investigate the protective effect of royal jelly in varicocele-induced oxidative damages and disrupts spermatogenesis cycle of rat testis by biochemical and histopathological methods. The present study demonstrates, for the first time, the possible effects of royal jelly on reducing the complications of experimental varicocele in rats.

2. Material and methods

2.1. Animals and drugs

Twenty-one adult male Wistar albino rats (aged 7–8 weeks) weighting 200 ± 50 g were randomly divided into three groups (Carmignani et al., 1983; Sofikitis et al., 1992; De Stefani et al., 2005; Amirshahi and Najaﬁ, 2013). Group I: Control (healthy rats without induction of varicocele).

Group II: varicocele induction and injection of physiological saline daily with 3 mM solution via subcutaneous injection.

Group III: varicocele induction and treatment with RJ (daily with 200 mg/kg) via gavage directly into the stomach (Nekeety et al., 2007; Amirshahi and Najaﬁ, 2013; Zamatkesh et al., 2014; Ahmadnia et al., 2015).

All animal experiments comply with the ARRIVE guidelines and carried out in accordance with the U.K. Animals (Scientific Procedures) Act, 1986 and associated guidelines, EU Directive 2010/63/EU for animal experiments, and were approved by the Ethics Committee of Lorestan University of Medical Sciences.

2.2. Experimental procedure

For induction of varicocele, the animals were anaesthetized with Ketamine (Razak, Iran), IP = 60 mg/kg and Xylazine (Triratn, Germany), 1 mg/kg. A vertical midline section with a length of 3–4 mm was created at the abdominal region. After detection of the left renal vein and the insertion point of the internal spermatic vein, 50% of the left ventricular vein was blocked by 20 gauge angio catheter and 4-0 silk suture. The surgery site was closed with 3-0 silk suture (Ozturk, Koca et al., 2013). All of the surgical procedures were performed under sterile conditions (Köksal, Erdörü et al., 2002; Bernal, Baldwin et al., 2009). The animals were housed in cages (three or four animals/cage) under controlled temperature (22 ± 2 °C). They had free access to standard diet, tap water and 12 h of light and darkness for five weeks. In this research, the ethical procedures used in animal experimentation were applied. At the end of the experiment (6 weeks after creation of varicocele), all the animals were sacrificed. The abdominal cavity was opened up through a midline abdominal incision and the testes were excised quickly. Half of the testes were fixed with Bouin’s fixative (0.2% picric acid/2% (V/V) formaldehyde in PBS2% (V/V)) for histological evaluation (Najaﬁ et al., 2014). The other half was stored for sub-sequent biochemical assays at ~70 °C.

2.3. Antioxidant enzyme activity and MDA level

The testicular tissue samples thawed on ice and homogenized in PBS buffer by homogenizer. The protein content of the testicular tissue was assayed by the Bradford method (Bradford, 1976). This method relies on the attachment of Coomassie Brilliant Blue anionic forms to the arginine and lysine. Tissue catalase activity can be assayed using the procedure proposed by Aebi (Aebi, 1984). The basis of this method is to reduce the light absorbance of H2O2 at 240 nm. The reaction contained 0.1 ml of testicular homogenate, 1.9 ml 50 mM phosphate buffer, pH 7.0 and 1.0 ml of freshly prepared 30% (v/v) hydrogen peroxide (H2O2). Enzyme activity was expressed as unit’s /mg protein (Köksal et al., 2002; Kheradmand et al., 2009; Hanumanth Goud et al., 2012; Asad et al., 2018). SOD activity was evaluated by the method described by Rukmini (Rukmini et al., 2004). This method was based on the ability of the SOD to inhibit reduction of the nitro-blue tetrazolium. The reaction contained 2.7 ml of 0.067 M phosphate buffer, pH 7.8, ribo-flavin (0.05 ml of 0.12 mM), as well as nitro-blue tetrazolium (0.1 ml of 1.5 mM), methionine (0.05 ml of 0.01 M) and 0.1 ml of enzyme samples. One unit of SOD is defined as the amount of enzyme required to inhibit the reduction of NBT by 50% under specific conditions. The absorbance was measured at 560 nm. Results were expressed as units (U) of enzyme activity (u/mg protein). GPX was measured based on the Rotruck method (Rotruck et al., 1980). The samples were read at 420 nm and expressed as nmol/mg protein. The reaction contained Tris-HCl buffer (2.0 ml of 0.4 M), pH7.0, sodium azide (0.01 ml of 10 mM), enzyme (0.2 ml), glutathione (0.2 ml) and H2O2 (0.5 ml of 0.2 mM). The contents were incubated at 37 °C for 10 min. Then, the reaction was terminated by addition of 0.4 ml 10% (v/v) TCA, and was centrifuged at 5000 rpm for five minutes. MDA test was performed using thiobarbi-turic acid reaction. This method was described as the Esterbauer method (Esterbauer and Zollern, 1989). MDA reacts with thiobarbituric acid to give a red compound that is absorbed at 532 nm in a spectro-photometer (Esterbauer and Cheeseman, 1990; Sohrabipour et al., 2013; Najaﬁ et al., 2014). The absorption level was obtained using the molar coefficient, based on μmol/mg of protein.
μ was placed in an incubator (37 °C with 5% CO2) for 25 min. To count Epididymis caudal was minced with scissors to release the sperm and medium (Hams F10) containing 0.5% bovine serum albumin. separated from the testis and placed in a Petri dish containing 2 ml of

<table>
<thead>
<tr>
<th>Score</th>
<th>Histological criteria</th>
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<tbody>
<tr>
<td>10</td>
<td>Full spermatogenesis and perfect tubules</td>
</tr>
<tr>
<td>9</td>
<td>Disorganized spermatogenesis; slightly impaired spermatogenesis</td>
</tr>
<tr>
<td>8</td>
<td>A few spermatooza present</td>
</tr>
<tr>
<td>7</td>
<td>No spermatooza but many spermatids present</td>
</tr>
<tr>
<td>6</td>
<td>No spermatooza, only a few spermatids present</td>
</tr>
<tr>
<td>5</td>
<td>No spermatooza or spermatids but many spermatocytes present</td>
</tr>
<tr>
<td>4</td>
<td>Only a few spermatocytes present</td>
</tr>
<tr>
<td>3</td>
<td>No spermatocytes only spermatogonia present</td>
</tr>
<tr>
<td>2</td>
<td>No germ cells but only Sertoli cells present</td>
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<tr>
<td>1</td>
<td>No germ cells and no Sertoli cells present</td>
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2.5. Sperm characteristics

To evaluate sperm parameters, the Cauda epididymis was carefully separated from the testis and placed in a Petri dish containing 2 ml of medium (Hams F10) containing 0.5% bovine serum albumin. Epididymis caudal was minced with scissors to release the sperm and was placed in an incubator (37 °C with 5% CO2) for 25 min. To count the sperm, the semen was diluted with 1:2 proportion. Approximately, 15 μl of the diluted sperm suspension was transferred to each counting chamber of the hemocytometer and all the squares (1–9) were counted. Sperm count and viability were done under a light microscope. Total numbers of motile sperms per 100 sperms were counted. Percentage of sperm viability was accessed by the one step eosin-nigrosine staining technique. Non stained cells were considered as alive and in dead cells, stain had passed through the membrane and colored as orange-red. The sperm heads were counted and expressed as million/ml of suspension.

2.6. Data analysis

The statistical analysis was carried out using the SPSS package, version 16.0. The Kolmogorov-Smirnov and the Shapiro-Wilk’s test was used to check the normal distribution of data. The differences among the groups were assessed by the analysis of variance (ANOVA) followed by the Duncan test. The results expressed as mean ± (S.E.M.) and values of P < 0.05 were considered statistically significant.

3. Results

3.1. Antioxidant enzyme activity and MDA level

Antioxidants enzyme activity, MDA levels and total antioxidant capacity (TAC) in the three groups are shown in Fig. 1 and 2. Our results showed, untreated varicoceleized model (Group II) had a significantly (p < 0.05) decrease in CAT (0.108 ± 0.0178), SOD (0.160 ± 0.002), GPx (8.425 ± 0.149) activities and TAC (1.16 ± 0.15). These results indicate that the administration of RJ in group III significantly (p < 0.05) increased activity of CAT (0.223 ± 0.0178), SOD (0.177 ± 0.002), GPx (9.575 ± 0.149) and TAC (2.67 ± 0.15). The activities of these antioxidant enzymes in RJ Group were roughly similar to that of the control values.

Our findings suggest that induction of varicocele can significantly (P < 0.05) increased MDA level (4.055 ± 0.219) compared to the control group. In this regard, the administration of royal jelly in group III significantly (P < 0.05) reduced MDA levels (1.960 ± 0.219) compared to group II (Fig. 2).

3.2. Histopathology

Our finding suggests that in the control Group (I) the testes had normal morphologic characteristics. The seminiferous tubules closely packed and basement membrane were regular (Fig. 3-A). The germ cells were organized in concentric layers when compared to the Group II and Group III testes (Fig. 3-B).

In Group II, there were various multifocal structural changes affecting the seminiferous tubules with loss of their normal architecture with decreased diameters. Widening of the spaces between the seminiferous tubules and atrophic cells in numerous seminiferous tubules can be seen (Fig. 4-A) (arrow). In this group, the basement membrane was thickened and necrosis tubular can be seen (Fig. 4-B). Also it was observed an increase in the histopathological damage score, tubular atrophy and decreases reduction in the the number of germ cells, compared to the control group (Fig. 4-C).

In Group III, the results showed a relatively normal developed histopathological feature (Fig. 5-A). In addition, the RJ exhibited relatively complete spermatogenesis, spermatooza and decrease in the luminal areas when compared to Group II. However, in some tubules, a decrease in the number of germ cells was observed (Fig. 5-B).

Results of the Johnsen score in Group I (control group) showed a significant (p < 0.05) increase in the mean score (9.5 ± 0.814) when compared to Group II and Group III. In Group II, there was a significant (p < 0.05) reduction in the mean score (6.04 ± 1.4) when compared to Group I and Group III. Our results showed that in the royal jelly group (Group III), the mean score (7.94 ± 1.5) significantly improved (p < 0.05) when compared to Group II (Fig. 6).

3.3. Sperm parameters

The results of the sperm parameters analysis are depicted in Fig. 7. In Group I, the sperm count and viability significantly increased viability were significantly higher (p < 0.05) when compared to Group II and Group III. This shows the deleterious effect of the varicocele on sperm quality. While the administration of 200 mg/kg RJ (III) significantly increased (p < 0.05) sperm viability in compared to the varicocele and saline group. In this experiment, RJ administration improved the sperm count. Although, these changes were not significant in compared to saline to the varicocele group (II).

4. Discussion

The negative effects of varicocele on fertility are widely recognized. The present study confirms previous studies that the induction of left varicocele in Wistar rats results in a bilateral increase in the level of MDA and statistically significant decrease in the level of antioxidant enzymes activities (p < 0.05) (Turner, 2001; Semercioz et al., 2003; Asadi et al., 2018). A variety of antioxidants have also been assessed for their ability to counteract oxidative stress caused by varicocele created by alternative mechanisms (Asadi et al., 2018). It seems that RJ protects the testicular tissue against oxidative stress in rats with varicoceles induction. We confirmed that rats with experimental varicocele and supplemented with 200 mg/kg/day of RJ (III) had a significant increase in the antioxidant enzyme activity and testicular content of SOD, CAT, GPx and TAC compared to the group II. These observations are in

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accordance with the outcomes obtained by previous researchers (Nekeety et al., 2007; Hassan, 2009; Silici et al., 2011; Najaﬁ et al., 2014; Zahmatkesh et al., 2014). The three primary antioxidant enzymes, namely CAT, SOD, GPx and TAC contained in the cells were thought to be necessary for life in all oxygen metabolizing cells (Varjovi et al., 2015). The positive effects of RJ antioxidant activities in testes, liver, kidneys, Pancreas, tumor cells, sexual efficiency in adult male rats and sub fertile men are widely recognized (Nekeety et al., 2007; Amirshahi and Najafi, 2013; Shirzad et al., 2013; Ahmadnia et al., 2015; Nejati et al., 2016). Antioxidant property related to the function of the proteins of RJ caused inhibition of free radical activity (Nagai and Inoue, 2004). More than 29 antioxidant bio peptides were purified from the hydrolysis of RJ proteins. Several mechanisms for the antioxidant properties of RJ have been identified including hydroxyl radical scavenging activity, hydrogen peroxide scavenging activity, metal-chelating activity and superoxide radical scavenging activity (Guo et al., 2009).

The present study indicated that lipid peroxidation (MDA) in testis significantly increased in varicocelized rats. While RJ treatment significantly ameliorated the decrease in MDA levels (Decrease by about 50%) in varicocelized rats compared to the group II. This result agrees with previous studies which have demonstrated the RJ caused statistically significant decreases in the level of MDA of testis (Hassan, 2009; Najafi et al., 2014; Zahmatkesh et al., 2014), liver and kidney (Cavusoglu et al., 2009; Karadeniz et al., 2011; Silici et al., 2011). Malondialdehyde level is commonly known as a marker of oxidative stress and the antioxidant status in patients with varicocele (Gawel et al., 2004). One of the mechanisms associated with decrease MDA level is related to the vitamins in the royal jelly. The royal jelly contains vitamin C, vitamin E and arginine. Vitamin E and C is a well-documented antioxidant and has been shown to inhibit free-radical induced damage to sensitive cell membranes of the testis and reduced lipid peroxidation in tissue estimation by malondialdehyde (Hassan, 2009). Moreover, vitamin E had a significantly increased activity level of SOD, CAT and GPx and a reduced lipid peroxidation which is evidenced by significant reduction in level of MDA (Saalu et al., 2013). Also in 2009, Guo confirmed that royal jelly proteins (RJPs) hydrolyzed with protease
N show the strong antioxidative activity against the peroxidation of linoleic acid (Guo et al. 2009).

Furthermore, on microscopic examination, histopathologic changes were observed in varicocelesed rats. These changes included decrease in tubular diameter, spermatogenic arrest, disorganization, degeneration and decrease in the number of germ cells. While our histological findings showed that RJ administration caused less degenerative alterations in rats with varicoceles induction. These results confirmed the results of other researchers (Karadeniz et al., 2011). It seems that one of the effective factors that lead to smaller histological modifications is the increase in anti-oxidant. Increasing activities of these antioxidant enzymes in the RJ group implied a decrease in the histopathologic changes after varicocele induction reflects the important role that these enzymes play in scavenging free radicals. Our findings are similar to results of other investigators such as Najafi et al. (Najafi et al., 2014), Zahmatkesh (Zahmatkesh et al., 2014) and Hassan (Hassan, 2009) for testis tissues in which RJ administration caused increase SOD, CAT, GPX and TAC levels. Other reports showed that treatment of diabetic rats with RJ significantly increased CAT activity and improved histological changes in kidney and testis tissues (Ghanbari et al., 2015). Moreover, it was demonstrated that the protein fractions in RJ have high antioxidative activity and scavenging ability against free radicals such as superoxide anion radical, DPPH (1,1- diphenyl-2-picrylhydrazyl) radical, and hydroxyl radical (Nagai and Inoue, 2004; Zahmatkesh et al., 2014).

In the current study, we showed that varicocelesed rats had...
significantly low levels of sperm count and viability compared with the control group. While, treatment with RJ resulted in improvement in sperm count and significant increase in sperm viability. These results are in agreement with the findings of previous studies (Nekeety et al., 2007; Sanafi et al., 2007; Hassan, 2009; Zahmatkesh et al., 2014; Jalali et al., 2015). One mechanism is that the RJ enhances production of the seminal fluid from the secondary sex organs, which play the main role in sperm’s viability and motility (Sanafi et al., 2007). Also, RJ contains acetylcholine, L-arginine and Carnitine amino acids, which helps to stimulate gonadotropin secretion of the hypothalamic level and essential for spermatogenesis (Mitsushima et al., 2008). Moreover the amino acid and fatty acids specific to RJ, especially 10-hydroxy-2-decenoic acid content of RJ may play a role as well by enhancing acrosome reaction, sperm count, sperm viability and improving fertilization (Renard et al., 1996; Comhaire et al., 2000). Another mechanism is that the zinc supplement in royal jelly can increase testosterone levels and increase fertility. Studies confirmed that testosterone is essential for spermatogenesis from spermatogonia to spermatids (Boselli et al., 2003). In the current study, although the sperm count in group III was higher compared with that in groups II, the difference was not significant. It seems that the sample size, effective dose and short-term of RJ administration might have been responsible for the lack of significant difference.

5. Conclusion

Varicocele has negative effect on male reproductive system. However, Royal jelly protects rat testis against oxidative stress effects of varicocele. This protection may be due to the enhancement of antioxidant enzymes formation and the suppressive effects of lipid peroxidation and free radical generation. However, further experimentation with higher dosage and long-term for investigating the association

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**Fig. 5.** Histopathological sections of rat testes in the RJ group.
A-Architecture of the seminiferous tubules was relatively normal (H-E, X100).
B-seminiferous tubules appeared with regular outlines and normal lining of the stratified germinal epithelium (H-E, X200).

**Fig. 6.** Effect of royal jelly on Johnsen score following varicocele induction in Rat.
The values are expressed as mean ± SEM (n = 7). P values < 0.05 were considered significant.
a, b, c Mean values with the same letter do not differ statistically.

**Fig. 7.** Effect of royal jelly on sperm parameters following varicocele induction in Rat.
The values are expressed as mean ± SEM (n = 7). P values < 0.05 were considered significant.
a, b, c Mean values with the same letter do not differ statistically.
between AJB administration and sperm parameters is warranted.

Notes

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Conflict of interest

The authors declare that they have no conflicts of interest.

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