



KIRs gene content diversity in Iranians with urothelial bladder cancer

Elham Jamali² · Shaghik Barani¹ · Fahimeh Yousefinejad² · Ali Ariaifar³ · Ghoalm Reza Talei⁴ · Abbas Ghaderi^{1,2}

Received: 28 February 2018 / Accepted: 4 June 2018 / Published online: 2 July 2018
© Springer Nature B.V. 2018

Abstract

Natural killer cells (NK) are the first arm of the innate immune system in defense against tumor and infection. 16 distinct Killer-cell immunoglobulin-like receptors (*KIRs*) are involved in orchestrating NK cell function. The *KIR* family contains 14 genes and 2 pseudogenes. Six of these receptors are activating (a*KIR*) and the remaining receptors are inhibitory *KIRs* (i*KIR*), that interact with MHC-I molecules; producing signals which stop NK cell function. In the current study, we have investigated the genomic diversity of *KIRs* and determining the A and B haplotypes as well as Bx subsets in 119 patients with bladder cancer and 200 healthy controls to find out if there is an association between *KIR* system and susceptibility to bladder cancer. Polymerase chain reaction with sequence specific primers (SSP–PCR) typing system was used to determine the *KIR* gene profile. The results implicated decreased frequency of inhibitory *KIR2DL2* and activating *KIR2DS2* while increased frequency of CxT4 genotypes in patients compared with healthy controls. Among Bx subsets, the CxT4 gene cluster is more frequent in bladder cancer patients compared to controls. Our results provide a conclusion that *KIR2S2* and *KIR2L2* may play a protective role against bladder cancer development while the CxT4 gene cluster may underlie susceptibility to bladder cancer in Iranian population.

Keywords Killer immunoglobulin like receptors (*KIRs*) · Bladder cancer · Natural killer cell · Polymerase chain reaction with sequence specific primers (PCR–SSP)

Introduction

Urothelial bladder cancer (UBC) is the most common malignancy of urinary system [1, 2]. It has high morbidity and mortality rate, as well as high rate of recurrence to invasive forms [2]. The disease incidence in men is three times more than women. A combination of environmental and genetic factors affect susceptibility to UBC. Tobacco smoking, age, and occupational exposure to polycyclic hydrocarbons like aromatic amines are most important risk factors for UBC [3–5]. Along with the development of the molecular

immunology, the association of immune response genes and various diseases has been highlighted. Human leukocyte antigen (HLA) and Killer-cell immunoglobulin-like receptor (*KIR*) genotyping studies has been a field of interest; is being extensively typed in many diseases [6, 7].

KIRs are one of the immune response genes which play crucial role in NK cell mediated functions. *KIR* molecules are a polymorphic family of receptors encoded by a gene cluster on 19 q13.4. 14 distinct *KIR* genes and 2 pseudogenes have been identified. The *KIR2DL1*, *2DL2*, *2DL3*, *2DL5*, *3DL1*, *3DL2*, and *3DL3* have an inhibitory function; containing immunoreceptor tyrosine-based inhibitory motif (ITIM) in their cytoplasmic tail. *KIR2DS1*, *2DS2*, *2DS3*, *2DS4*, *2DS5*, and *3DS1* are activating *KIRs*. *2DP1* and *3DP1* are known as pseudogenes [7]. The net balance of signal from activating and inhibitory receptors will determine the NK cell final destiny. According to *KIR* gene content two distinct haplotypes has been introduced. Haplotype A containing *KIR2DL3*, *2DP1*, *2DL1*, *3DL1*, *2DS4*, and haplotype B which is more variable and includes diverse combinations of *KIR2DS1*, *2DS2*, *2DS3*, *2DS5*, *2DL2*, *2DL5*, and *3DS1*. *KIR2DL4*, *3DL2*, *3DL3*, and *3DP1* are as framework genes

✉ Abbas Ghaderi
ghaderia@sums.ac.ir

¹ Shiraz Institute for Cancer Research, School of Medicine, Shiraz University of Medical Sciences, Shiraz, Iran
² Department of Immunology, School of Medicine, Shiraz University of Medical Sciences, Shiraz, Iran
³ Department of Urology, School of Medicine, Shiraz University of Medical Sciences, Shiraz, Iran
⁴ Hepatitis Research Center, Lorestan University of Medical Sciences, Khoramabad, Iran

which present on both haplotypes. Inhibitory *KIRs* recognize HLA I molecules. *KIR2DL2* and *KIR2DL3* recognize HLAC1 (HLAC with Asp at position 80), *KIR2DL1* bind HLAC2 (HLAC with Lys at position 80), *KIR3DL2* ligates with HLAA3/A11, and *KIR3DL1* binds HLABw4 (HLA B allotype containing Bw4 epitope). The ligands for activating *KIR* receptors are mostly unknown, but some of them can recognize HLA class I with strongly lower affinity than inhibitory ones [8–10].

Due to different haplotype inheritance patterns of *KIR* from parents, as well as the allelic polymorphism of *KIR* genes, the *KIR* gene cluster indicates extensive diversity. This diversity may affect susceptibility or resistance to different diseases. *KIR* genotyping studies have been done in autoimmune diseases, cancers, hematologic malignancies, etc. [11]. Such studies has not been extensively done in UBC patients. According to our information, only one *KIR* genotyping study with limited sample size ($n=65$) has been done by Middleton et al. on UBC patients. According to this study *KIR2DS4* and *KIR3DL1* are more frequent in UBC patients than healthy normal controls [12].

The aim of this study is to find out the diversity of *KIR* genes, genotypes and haplotypes and their comparison between UBC patients and healthy controls.

Material and method

Study subjects

119 patients with pathological confirmed bladder cancer (mean age: 65.23, 83% male and about 17% female) and 200 healthy controls were included in this study. The cases were recruited at Namazi Hospital and controls from Motahari Clinic. Patients and healthy controls were matched by age (± 5 years) and sex (demographic statistics are shown in Table 1). Both groups were ethnically matched and were from the Fars province in the southern part of Iran. Furthermore, bladder cancer patients were categorized to advanced and early stage, according to the TNM staging system. The study was also reviewed and approved by the Medical Research Ethics Committee of Shiraz University of Medical Sciences (EC-9372-7202).

Table 1 Demographic characteristics of cases and controls

Group	Bladder cancer n = 119	Controls n = 200
Mean of age \pm SD	65.23 \pm 21.10	61.6 \pm 14.03
%Female (N)	16.8 (20)	19.0 (38)
%Male (N)	83.1 (99)	81.0 (162)

KIR genotyping

Genomic DNA from all the subjects was extracted from whole blood samples using a QIAamp DNA Mini Kit (Qiagen, Germany). PCR-SSP method was used to characterize the presence/absence of 16 *KIR* genes in UBC patients and healthy controls. 100 ng of genomic DNA was amplified in 7.5 μ l Taq DNA polymerase 1 \times Master Mix RED. Specific primers designed by Vilches et al. were used to detect specific *KIR* genes [13] and primers designed by Ashouri et al. [14] for segregation of *KIR2DS4* full and deleted variants. In addition to specific primers three pair of internal controls were used to amplify 16 *KIR* genes in range of 0.4–2.6 μ M in specific primers and 0.1 μ M in internal controls [13].

PCR reactions were performed in the ABI thermal cycler according the following thermal condition: initial denaturation for 2 min at 95 $^{\circ}$ C, then 10 cycles genes of 10 s at 94 $^{\circ}$ C and 40 s at 65 $^{\circ}$ C; and 20 cycles of 20 s at 94 $^{\circ}$ C, 20 s at 61 $^{\circ}$ C and 30 s at 72 $^{\circ}$ C for final extension [13]. UCLA reference DNA with known *KIR* genotype, provided by Professor Rajalingam and co-workers was used to check the accuracy and specificity of the method [14]. *KIR* typing was done by detecting known PCR product size by comparing with DNA ladder (50 bp). Finally, we typed 11 unique genotypes again to validate the accuracy of data.

Data analysis and statistical methods

We determined the frequency of each *KIR* gene, A and B haplotypes, Bx subtypes as well as *KIR* gene clusters and compared them between patients and control group by two-tailed Chi square test of association in *Graphpad prism software* and $p < 0.05$ was considered to be statistically significant. The significant results were adjusted by Yates' correction. The *KIR* haplotypes based on gene content divided to Group A haplotypes with specified gene contents (*KIR3DL3-2DL3-2DP1-2DL1-3DP1-2DL4-3DL1-2DS4-3DL2*) and group B haplotypes with variable gene contents (*KIR2DL2-2DL5-2DS1-2DS2-2DS3-2DS5-3DS1*) which did not occur in A haplotype [15]. In the *KIR* gene complex two frequently occurring gene clusters are introduced due to the linkage disequilibrium, the C4 cluster with *KIR2DS2-2DL2-2DS3-2DL5* genes is located at the centromeric half and T4 cluster with *KIR3DS1-2DL5-2DS1-2DS5* genes is located at the telomeric half of the complex. So the Bx genotype will be divided into four subtypes C4Tx, CxT4, C4T4, and CxTx.

Results

Comparison of *KIR* genes frequency in bladder cancer patients and healthy controls

To determine the *KIRs* role as a genetic risk for bladder cancer, we studied the presence/absence of 16 different *KIR* genes and *KIR2DS4* variants in 119 bladder cancer patients and 200 healthy controls. The distribution of *KIR* genes in patients with bladder cancer and healthy controls is illustrated in Table 2. The framework genes *KIR2DLA*, *3DL2*, *3DL3* and *3DP1* were present in all 319 individuals.

We observed that A haplotypes associated *KIR* genes (*2DL1* *2DL3* *3DL1* *2DS4*) had higher frequency (> 89.5%) than B haplotypes associated ones both in cases and controls. *KIR2DL1* (99.1%) and *KIR2DS4* (91.5%) were the highest while the *KIR2DL2* (53.7%) and *KIR2DS* (36.9%) were the lowest inhibitory and activating *KIR* genes respectively in bladder cancer patients. We found a highly significant decrease in the carrier frequency of the inhibitory *KIR2DL2* (53 vs. 69% p value: 0.00690, OR 1.9, CI 1.2–3.1) and activating *KIR2DS2* (52 vs. 69%, p value: 0.0037, OR 2.046, CI 1.2–3.2) in bladder cancer patients compared to controls (Table 2).

To comparison the variants of *KIR2DS4* between cases and controls, the individuals were categorized into three groups: (A) Del: individuals who had only the deletion variant of *KIR2DS4* (B) Full: individuals who had only the full-length *KIR2DS4*, and (C) Full/Dell: individuals who had both full-length and deletion variant of *KIR2DS4* together. However we found an increase in the carrier frequency of *KIR2DS4* Del in bladder cancer (76.1%) in comparison to CNs (72.0%) but there was no significant difference (Table 3).

Comparison of *KIR* haplotype frequency in bladder cancer patients and healthy controls

The frequency of AA genotype carriers was higher (26.8 vs. 21.5%) while the Bx genotype carrier frequency was lower (73.2 vs. 78.5) in patients with bladder cancer compared with healthy controls, however there were no significant difference between them. Furthermore, the difference between the frequency of A and B haplogroup between patients and control group was not statistically significant (data are shown in Table 4).

Table 2 Frequency of *KIR* genes in bladder cancer patients and healthy controls

KIR gene	Bladder cancer n = 119		Controls n = 200		Bladder cancer versus controls		
	%F	(N+)	%F	(N+)	P value	Odds ratio	95% CI
A haplotypes associated <i>KIR</i> genes							
<i>2DL1</i>	99.1	(118)	99.5	(199)	> 0.9	0.59	0.03–11.3
<i>2DL3</i>	93.2	(111)	89.5	(179)	0.3	1.62	0.71–3.5
<i>3DL1</i>	91.5	(109)	96.0	(192)	0.130	0.45	0.17–1.22
<i>2DS4</i>	91.5	(109)	96.5	(193)	0.070	0.39	0.15–1.11
B haplotypes associated <i>KIR</i> genes							
<i>2DL2</i>	53.7	(64)	69.5	(139)	0.0069*	1.9	1.2–3.1
<i>2DL5</i>	62.1	(74)	61.0	(122)	0.9	1.05	0.65–1.67
<i>3DS1</i>	42.0	(50)	33.5	(67)	0.14	1.43	0.9–2.2
<i>2DS1</i>	42.0	(50)	43.5	(87)	0.81	0.94	0.6–1.49
<i>2DS2</i>	52.1	(62)	69.0	(138)	0.0037*	2.046	1.2–3.2
<i>2DS3</i>	42.0	(50)	49.5	(99)	0.2	0.73	0.47–1.16
<i>2DS5</i>	36.9	(44)	29.0	(58)	0.17	1.43	0.87–2.32
Framework genes/pseudogenes							
<i>2DL4</i>	100	(119)	100	(200)			
<i>3DL2</i>	100	(119)	100	(200)			
<i>3DL3</i>	100	(119)	100	(200)			
<i>2DP1</i>	99	(118)	99.5	(199)	> 0.9	0.59	0.03–11.3
<i>3DP1</i>	100	(119)	100	(200)			

Frequency of *KIR* genes expressed as percentage and defined as the number of individuals with *KIR* genes (N+) divide by number of individuals studies in the given study group (n). Con controls, (*) significant p value Yates correction

Comparison of carrier frequency of Bx subtypes in bladder cancer patients and healthy controls

The frequency of Bx subtypes was determined and the analysis of the results indicates that the CxT4 genotypes carriers was significantly higher in patients compared to controls (20 vs. 8.5%; p-value: 0.004, OR 2.76, CI 1.43–5.44). In addition, C4 gene cluster containing *KIR2DS2-2DL2-2DS3-2DL5* is more frequent in the control group compared to controls (p-value: 0.03, OR 0.59,

CI 0.36–0.94). The distribution of Bx subtypes in patients with bladder cancer and healthy controls is illustrated in Table 5.

KIR genotypes diversity in the study population

34 distinct genotypes were found among 319 individuals, which is illustrated in Table 6. These genotypes were defined by referring to the allele frequency database (<http://www.allelefreqencies.net>). We found 11 unique genotypes which

Table 3 Distribution of *KIR2DS4* variants in bladder cancer patients and healthy controls

KIR2DS4 variants	Bladder cancer n = 109		Control n = 193		Bladder cancer versus controls		
	%F	(N)	%F	(N)	p Value	Odds ratio	95% CI
Del	76.1	(83)	72.0	(139)	0.49	1.24	0.71–2.09
Full	16.5	(18)	22.2	(43)	0.29	0.69	0.38–1.24
Full/Dell	6.4	(7)	5.1	(10)	0.79	1.25	0.48–3.54
Missing	0.9	(1)	0.4	(1)			

The number (n) that is exhibited below each group is the number of people who were 2DS4 positive in each group that is categorized to three types (Deletion, Full and Full/Deletion) of *2DS4* gene. Two tailed Fisher exact probability (p) test, odd ratio with 95% CI was calculated by Graph pad prism software and p < 0.05 was considered to be statistically significant

Table 4 Frequency of KIR genotypes and haplotypes in Bladder cancer patients and healthy controls

	Bladder cancer n = 119		Controls n = 200		Bladder cancer versus controls		
	%F	(N+)	%F	(N+)	p Value	Odds ratio	95% CI
Genotype							
AA	26.8	(32)	21.5	(43)	0.27	1.34	0.79–2.29
Bx	73.2	(87)	78.5	(157)			
Haplotype							
A	55.8	(133)	53.9	(217)	0.43	0.74	0.35–1.56
B	44.1	(105)	46.0	(185)	0.21	0.61	0.28–1.30

Frequency of KIR genotypes/haplotypes presented as percentage and defined as the number of individuals with genotype/haplotype (N+) divided by number of individuals studies in the given study group (n). Con controls

Table 5 Frequency of Bx subtypes in bladder cancer patients and healthy controls

KIR genotypes/clusters	Bladder cancer n = 119		Controls n = 200		Bladder cancer versus controls		
	%F	(N+)	%F	(N+)	p value	Odds ratio	95% CI
C4Tx genotype	21.0	(25)	33.0	(66)	0.052	0.55	0.31–0.96
CxT4 genotype	20.0	(24)	8.5	(19)	0.0114*	2.76	1.43–5.44
C4T4 genotype	14.2	(17)	15.0	(30)	> 0.9	1.02	0.51–1.09
CxTx genotype	17.6	(21)	21.0	(42)	0.7600	0.87	0.47–1.57
C4 gene cluster	35.2	(42)	48.0	(96)	0.0359*	0.59	0.36–0.94
T4 gene cluster	34.4	(41)	24.5	(49)	0.07	1.62	0.99–2.62

Two tailed Fisher exact probability (p) test was calculated by Graph pad prism software and *P < 0.05: statistically significant; based on two-tailed Chi square with Yates' correction

No significant P value was found

Table 6 KIR genotypes distribution in the study population

# Genotype	KIR ID	Group	3DL1	2DL1	2DL3	2DS4	2DL2	2DL5	3DS1	2DS1	2DS2	2DS3	2DS5	2DL4	3DL2	3DL3	2DP1	3DP1	Bladder cancer n=119		Control n=200			
																			F%	(N)	F%	(N)		
1	1	AA																			0.5	1		
2	14	Bx																		26	31	21	42	
3	23																					0.5	1	
4	200																				0.8	1		
5	2																						0.5	1
6	27																						0.5	1
7	8																				4.0	5	4	8
8	10																						3.0	6
9	12																				1.6	2	1	2
10	19																						1	2
11	18																				3.2	4	1	2
12	58																						0.5	1
13	4																				4.8	6	1	2
14	31																						0.5	1
15	9																						1	2
16	3																				0.8	1	1	2
17	5																				0.8	1		
18	13																				8.0	10	4	8
19	11																				2.4	3		
20	7																						1	2
21	382																				1.6	2		
22	6																				2.4	3		
23	144																						0.5	1
24	190																				8.8	11	15	30
25	81																				0.8	1		
26	297																				0.8	1	1.5	3
27	180																				5.6	7	3.5	7
28	317																				9.6	12	11	22
29	71																						5.5	11
30	113																						2.5	5
31	90																						0.5	1
32	73																				4	5	5.5	11
33	308																				8.8	11	12.5	25
34	69																				1.6	2		

The presence and absence of a KIR gene are indicated by a shaded and white box, respectively. KIR ID assigned by the Allele Frequency Net Database

only present in one individual and were confirmed with sets of primers designed by Vilches et al. [13] and Ashouri et al. [14].

Discussion

KIR–HLA interactions determine the diversity of NK cell function; so that different combinations of KIR–HLA may affect susceptibility or resistance to some diseases [16]. According to our results, *KIR2DL2* and its activating

counterpart; *KIR2DS2* are less frequent in UBC patients compared with controls [17].

KIR2DL2 and *KIR2DL3* are inhibitory receptors which interact with *HLA-C1* and induce the inhibitory signals that dampen NK cell function; the *KIR2DL2* ligation is much stronger because of polymorphisms at two positions in ligand-binding site [18]. In accordance with our result, the protective role of *KIR2DL2* was demonstrated in patients with breast cancer, kidney disease and patients infected by HIV-1 [19] while its predisposing role was reported in leukemia [20].

KIR2DS2 which can bind *HLA-C1* with lower affinity than *KIR2DL2*, is associated with inflammatory condition, increased amount of IFN- γ secretion in patients infected by HIV-1 [19] and some autoimmune disorders [21] such as type 1 diabetes, rheumatoid arthritis, vasculitis and scleroderma [22]. Similar to our data, the frequency of *KIR2DS2* was lower in patients with HCV infection while in contrast with our result, its presence has been shown a protective role against colorectal cancer [23].

Decrease in the presence of *KIR2DS2* and *KIR2DL2* would imply that the most of UBC patients are probably have A haplotype than B. We observed a non-significant increase in AA genotypes in these patients than controls, while a lower frequency of AA genotype was reported in Iranians with head and neck squamous cell carcinomas than controls [24].

It is presumable that the *KIR2DS2* and *KIR2DL2* absence, result in more inhibition of NK cells in UBC patients. Since *KIR2DL2* are in strong linkage disequilibrium with *KIR2DS2*, lower frequencies of the carriers of both genes in UBC patients compared to controls remains to be studied in further studies as well as along with other candidate risk related genes in bladder cancer.

Considering the lower affinity of *KIR2DL2* than *KIR2DL3* for *HLA-C1*, the presence of *KIR2DL3* in the absence of *KIR2DL2*, due to decrease in the inhibitory signals and as a result, the chronic activation in NK cell and induction of inflammation in tumor microenvironment [16] which is linked to the development and progression of many cancers [25, 26]. Furthermore, lack of *KIR2DS2* may be decrease the NK cell's activity and possibly hampers the immunosurveillance. Momot et al. introduced some rare genotypes which had *KIR2DS2* but lacking *KIR2DL2* and were associated to susceptibility to scleroderma. It has been shown that *KIR2DS2* in the absence of *HLA-C1* can favor to NK cell activation [27].

Furthermore, our results show that CxT4 genotype which contains more activating receptors is significantly more frequent in UBC patients which is consistent with our previous research findings that the frequency of CxT4 genotype carriers was higher in patients with head and neck, laryngeal and oral cavity squamous cell carcinomas compared with healthy

controls [24]. It is assumed that the absence of inhibitory *KIR2DL2* in CxT4 cluster along with unknown tumoral antigenic stimuli as well as chronic inflammatory context of infectious agent associated cancers such as bladder cancer may contribute to sustainable NK cell chronic activation that increases the likelihood of development of cancer through local inflammation. Study of *KIR* genes diversity in thyroid and cervical cancer patients indicates that the frequency of activating *KIR* genes is higher in patients compared to controls, a finding which is consistent with the current study and interpretation [28, 29].

Future studies of *KIR/HLA* genes diversity in patients with bladder cancer as well as, functional studies of NK cells lacking *KIR2DL2* and *KIR2DS2* would be valuable strategy to find out the effect of their simultaneous absence on the threshold of NK cell activation and NK cell meddling in cancer immunity.

Acknowledgements This study was supported by a grant from (Grant No: 93-7202) Shiraz University of Medical Sciences. We thank those who helped us to collect the samples. This study was part of an MSc dissertation in Shiraz Institute for Cancer Research.

Compliance with ethical standards

Conflict of interest The authors have declared no conflicting interest.

References

- Burger M, Catto JW, Dalbagni G, Grossman HB, Herr H, Karakiewicz P, Kassouf W, Kiemeny LA, La Vecchia C, Shariat S (2013) Epidemiology and risk factors of urothelial bladder cancer. *Eur Urol* 63(2):234–241
- Sexton WJ, Wiegand LR, Correa JJ, Politis C, Dickinson SI, Kang LC (2010) Bladder cancer: a review of non-muscle invasive disease. *Cancer Control* 17(4):256–268
- Jacobs BL, Lee CT, Montie JE (2010) Bladder cancer in 2010: how far have we come? *CA: Cancer J Clin* 60 (4):244–272
- Letašiová S, Medvedová A, Šovčíková A, Dušinská M, Volkovová K, Mosoiu C, Bartonová A (2012) Bladder cancer, a review of the environmental risk factors. *Environ Health* 11(S1):S11
- Chu H, Wang M, Zhang Z (2013) Bladder cancer epidemiology and genetic susceptibility. *J Biomed Res* 27(3):170
- Tiwari JL, Terasaki PI (2012) *HLA and disease associations*. Springer, New York
- Rajalingam R (2011) Human diversity of killer cell immunoglobulin-like receptors and disease. *Korean J Hematol* 46(4):216–228
- Cooley S, Trachtenberg E, Bergemann TL, Saetern K, Klein J, Le CT, Marsh SG, Guethlein LA, Parham P, Miller JS (2009) Donors with group B KIR haplotypes improve relapse-free survival after unrelated hematopoietic cell transplantation for acute myelogenous leukemia. *Blood* 113(3):726–732
- Manser AR, Weinhold S, Uhrberg M (2015) Human KIR repertoires: shaped by genetic diversity and evolution. *Immunol Rev* 267(1):178–196
- Thielens A, Vivier E, Romagné F (2012) NK cell MHC class I specific receptors (KIR): from biology to clinical intervention. *Curr Opin Immunol* 24(2):239–245

11. Kulkarni S, Martin MP, Carrington M (2008) The Yin and Yang of HLA and KIR in human disease. *Semin Immunol* 20(6):343–352
12. Middleton D, Vilchez J, Cabrera T, Meenagh A, Williams F, Halfpenny I, Maleno I, Ruiz-Cabello F, Lopez-Nevot M, Garrido F (2007) Analysis of KIR gene frequencies in HLA class I characterised bladder, colorectal and laryngeal tumours. *HLA* 69(3):220–226
13. Vilches C, Castano J, Gómez-Lozano N, Estefania E (2007) Facilitation of KIR genotyping by a PCR-SSP method that amplifies short DNA fragments. *HLA* 70(5):415–422
14. Ashouri E, Ghaderi A, Reed E, Rajalingam R (2009) A novel duplex SSP-PCR typing method for KIR gene profiling. *HLA* 74(1):62–67
15. Uhrberg M, Valiante NM, Shum BP, Shilling HG, Lienert-Weidenbach K, Corliss B, Tyran D, Lanier LL, Parham P (1997) Human diversity in killer cell inhibitory receptor genes. *Immunity* 7(6):753–763
16. Moesta AK, Parham P (2012) Diverse functionality among human NK cell receptors for the C1 epitope of HLA-C: KIR2DS2, KIR2DL2, and KIR2DL3. *Front Immunol*. <https://doi.org/10.3389/fimmu.2012.00336>
17. Pyo C-W, Guethlein LA, Vu Q, Wang R, Abi-Rached L, Norman PJ, Marsh SG, Miller JS, Parham P, Geraghty DE (2010) Different patterns of evolution in the centromeric and telomeric regions of group A and B haplotypes of the human killer cell Ig-like receptor locus. *PLoS ONE* 5(12):e15115
18. Moesta AK, Norman PJ, Yawata M, Yawata N, Gleimer M, Parham P (2008) Synergistic polymorphism at two positions distal to the ligand-binding site makes KIR2DL2 a stronger receptor for HLA-C than KIR2DL3. *J Immunol* 180(6):3969–3979
19. Lin Z, Kuroki K, Kuse N, Sun X, Akahoshi T, Qi Y, Chikata T, Naruto T, Koyanagi M, Murakoshi H (2016) HIV-1 control by NK cells via reduced interaction between KIR2DL2 and HLA-C* 12:02/C* 14: 03. *Cell Rep* 17(9):2210–2220
20. Augusto DG (2016) The impact of KIR polymorphism on the risk of developing cancer: not as strong as imagined? *Front Genet* 7:121
21. Aguilar AMO, Guethlein LA, Adams EJ, Abi-Rached L, Moesta AK, Parham P (2010) Coevolution of killer cell Ig-like receptors with HLA-C to become the major variable regulators of human NK cells. *J Immunol* 185(7):4238–4251
22. Miyashita R, Tsuchiya N, Yabe T, Kobayashi S, Hashimoto H, Ozaki S, Tokunaga K (2006) Association of killer cell immunoglobulin-like receptor genotypes with microscopic polyangiitis. *Arthritis Rheumatol* 54(3):992–997
23. Kim H-J, Choi H-B, Jang J-P, Baek I-C, Choi E-J, Park M, Kim T-G, Oh S-T (2014) HLA-Cw polymorphism and killer cell immunoglobulin-like receptor (KIR) gene analysis in Korean colorectal cancer patients. *Int J Surg* 12(8):815–820
24. Barani S, Khademi B, Ashouri E, Ghaderi A (2018) KIR2DS1, 2DS5, 3DS1 and KIR2DL5 are associated with the risk of head and neck squamous cell carcinoma in Iranians. *Hum Immunol*. <https://doi.org/10.1016/j.humimm.2018.01.012>
25. Janakiram NB, Rao CV (2014) The role of inflammation in colon cancer. *Inflamm Cancer*. https://doi.org/10.1007/978-3-0348-0837-8_2
26. Hagemann T, Balkwill F, Lawrence T (2007) Inflammation and cancer: a double-edged sword. *Cancer Cell* 12(4):300–301
27. Momot T, Koch S, Hunzelmann N, Krieg T, Ulbricht K, Schmidt R, Witte T (2004) Association of killer cell immunoglobulin-like receptors with scleroderma. *Arthritis Rheumatol* 50(5):1561–1565
28. Ashouri E, Dabbaghmanesh MH, Rowhanirad S, Bakhsh-ayeshkaram M, Omrani GR, Ghaderi A (2012) Activating KIR2DS5 receptor is a risk for thyroid cancer. *Hum Immunol* 73(10):1017–1022
29. Carrington M, Wang S, Martin MP, Gao X, Schiffman M, Cheng J, Herrero R, Rodriguez AC, Kurman R, Mortel R (2005) Hierarchy of resistance to cervical neoplasia mediated by combinations of killer immunoglobulin-like receptor and human leukocyte antigen loci. *J Exp Med* 201(7):1069–1075